Contribution to the species composition and taxonomic status of some *Rattus* inhabiting Southern Vietnam and Sundaland

Alexander E. Balakirev & Viatcheslav V. Rozhnov

ABSTRACT. The sequencing of Cyt b and COI mtDNA genes was used to investigate the species composition of Rattus rats in Southern Vietnam and establish an actual correspondence between revealed mtDNA haplogroups and taxonomical species for the group in the region. Seven specific level phylogroups corresponding to Rattus tanezumi, R. norvegicus, R. nitidus, R. andamanensis, R. exulans, R. argentiventer and R. tiomanicus were discovered, with European R. rattus apparently being absent. By comparing original genetic data with other sequences representing different localities in Indochina and the Sundaland regions from GenBank, transsundaic distribution of R. tiomanicus was shown in continental Indochina well beyond the Isthmus of Kra, along with the other species, R. argentiventer, overpassing the limit. The adequacy of species attribution and taxonomic names for three Indochinese species (R. andamanensis, R. argentiventer and R. tiomanicus) is discussed based on comparisons of morphological peculiarities and genetic data set for Vietnamese samples. Corrected nomenclature for specific level haplogroups/species of Rattus in Southeast Asia is proposed.

KEY WORDS: Rattus, Indochina, Vietnam, species composition, mammals, taxonomy.

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О видовом составе и таксономическом статусе некоторых форм *Rattus*, обитающих в Южном Вьетнаме и на Сундской Суше

А.Е. Балакирев, В.В. Рожнов

РЕЗЮМЕ. Для исследования фактического видового состава и определения таксономического соответствия, обнаруженных в регионе митохондриальных гаплогрупп представителей рода *Rattus*, обитающих на территории Южного Вьетнама, было проведено секвенирование генов цитохрома б (Cyt b) и цитохромоксидазы (COI) митохондриальной ДНК. Было выявлено семь филогрупп видового уровня, соответствующих видам *Rattus tanezumi*, *R. norvegicus*, *R. nitidus*, *R. andamanensis*, *R. exulans*, *R. argentiventer* и *R. tiomanicus*, при этом европейский *R.rattus* в регионе обнаружен не был. Сравнением оригинальных генетических данных с данными депонированными в GenBank показано транссундаическое распространение *R. tiomanicus*, который обнаружен в континентальном Индокитае (Южный Вьетнам) значительно севернее Перешейка Кра, равно как и *R. argentiventer*, который тоже переходит эту границу. На основании сравнения черт внешнего строения и молекулярных данных по выборке из Вьетнама обсуждается адекватность видового определения и применяемых видовых названий для трёх индокитайских видов *R. andamanensis*, *R. argentiventer* and *R. tiomanicus*. Предложена корректированная номенклатура для обозначения филогрупп видового уровня рода *Rattus* в Юго-Восточной Азии.

КЛЮЧЕВЫЕ СЛОВА: Rattus, Индокитай, Вьетнам, видовой состав, млекопитающие, таксономия.

Introduction

Species belonging to the genus *Rattus* are traditionally considered to be a group difficult to identify, especially in Southeast Asia and the Sunda Islands. The difficulty is caused by a similarity in general appearance, external morphology and skull structure between many of the species and their considerable geographical and ecological variability. This difficulty in identifi-

cation led to problems with species identification and sometimes gives rise to conflicting views on the identification of taxa in the fauna of the region. The current situation in the taxonomy of the group creates problems for ecologists, medical zoologists and other specialists in zoology and natural conservation.

During the last century, many studies were focused on investigating the biodiversity of the family Muridae in Southeast Asia. However, until recently Vietnam remained the least investigated area in the region in this respect. The taxonomy of the family Muridae was revised many times during this period (Ellerman, 1949; Ellerman & Morrison-Scott, 1966; Musser & Carleton, 1993, 2005; Nowak, 1999; Pavlinov, 2005) but until now could not be considered as finally established. The same is true for opinions on the correspondence of forms and species and their natural ranges and ecologies.

Different authors have significantly disagreed on the species composition of the fauna of continental Indochina. Van Peenen (1969) indicated seven species within the *Rattus* genus for the region (*norvegicus*, exulans, exiguus, rattus, slaideni, nitidus, and argentiventer). Musser (in Corbet & Hill, 1992), followed by Pavlinov (2005), in his most complete review of the mammals of Southeast Asia, stated the genus Rattus as also comprising seven species but in a different assemblage (norvegicus, exulans, rattus, nitidus, argentiventer, remotus, and osgoodi). Lunde & Nguyen Truong Son (2001) agreed with Musser's opinion on *Rattus* composition, with the addition of R. tanezumi considered as a separate species. Kuznetsov (2006) argued that the genus *Rattus* is even more complex and comprised 10 species at total (norvegicus, exulans, losea, rattus, koratensis, nitidus, argentiventer, remotus, osgoodi, and moliculus). If synonyms are taken into consideration, it follows that R. exiguous is a synonym of R. losea Swinhoe, 1870 and R. slideni is a synonym of R. tanezumi Temminck, 1844, which is in agreement with Musser & Carleton (1993) (or the synonym for *R*. koratensis in accordance with Kuznetsov, 2006), generally, it may be proposed the number of *Rattus* species in Indochina is in the limits from seven to ten.

A series of successful investigations of the genetic diversity in the genus in the Asia Pacific region (Chinen et al., 2005; Robins et al., 2007, 2010; Pages et al., 2010) were performed recently. Using the analysis of short mtDNA fragments (cytochrome b genes and subunit I cytochrome oxidase gene, Cyt $\,b$ and COI) (Ross et al., 2003; Robins et al., 2007), the possibilities to determine genetic species and estimate their proper taxonomic attributions was clearly demonstrated. A set of discrete specific genetic clades have been shown (Chinen et al., 2005; Robins et al., 2007), and minimal genetic distances to segregate specific levels taxa were estimated (Pages et al., 2010). Currently, ten specific level phylogroups have been characterized for Sundaland area, with more than 20 such groups recognized for entire Australasian region. The phylogenetic relationships among them are discussed in detail by Robins et al. (2007, 2010). More than 600 available sequences of Cyt b and COI genes in the mtDNA, the IRBP gene and some other genes of wild (non-laboratorial) representatives of Rattus were deposited in international gene databases (GenBank, NCBI, DDBJ) by December 31, 2010, providing plenty of material for an accurate definition of intrageneric taxonomy based on up-todate molecular approaches to actual natural biodiversity investigations. Nevertheless, the problem of reciprocal correspondence of the scientific species names for revealed species-level phylogroups with the generally accepted taxa names, initially described exclusively based on general appearance and some morphological traits in skull structure, is yet to be resolved for the general revision of the species composition in the genus.

Thus, in this paper, we endeavored to present some contribution to species composition of the genus *Rattus* from continental Indochina and Sundaland and estimate specific names usage for genetically delimitated species based on comparison of currently available molecular genetics and morphological data. Authors do not pretend for explicit taxonomic revision for the genus in the region, nevertheless the discussions listed bellow may be useful to shed some light to some important taxonomical questions and contribute to proper taxonomic names usage.

Materials and methods

Specimens studied

We analyzed field collections and other material obtained during a series of expeditions performed by the Joint Vietnam-Russian Tropical Research and Technological Centre from 2007 to 2011 in Vietnam. The materials were collected in Hoang Lien Nature Reserve (Lao Cai Province), Nam Cat Tien National Park (Dong Nai Province), Bu Gia Map Nature Reserve (Binh Phuc Province), Con Dao National Park (Ba Ria – Vung Tau Province), precincts of the Lo Go Xa Mat Nature Reserve (Tay Ninh Province), Binh Chau Nature Reserve near Xuyen Moc (Ba Ria - Vung Tau Province), Ngoc Linh Nature Reserve (Kon Tum Province), Gia Lai Province near Pleiku, Dak Lak Province near Ea Wy (Tay Nguyen Plateau) and in the rural areas surrounding Vinh Long, Hanoi, Hai Phong and Ho Chi Minh City. A map of the collection sites are shown in Fig. 1.

Preliminary species identification was based on field guides (Van Peenen, 1969; Lunde & Nguyen Truong Son, 2001) and on reviews by Corbet & Hill (1992) and Kuznetsov (2006). Captured rats were photographed, weighed and measured, after which a small piece of liver or muscle tissue was collected (fixed in 96% ethanol) for molecular investigations. We also collected the skull and skin for comparative morphological investigations, performed visually and using a stereomicroscope MBS-10. For preparing illustrations for the paper some of the skulls were photographed in more considerable details under macro mode by Olympus S-350 digital camera (8 Mpx). In general, we examined the morphological materials (skulls, skins and alcoholpreserved carcasses) for more than one hundred animals, most of which (72 individuals) were genotyped. The majority of the morphological materials are deposited at the Zoological Museum of Moscow State University, Moscow, Russia (ZMMU), and some specimens are deposited in the Zoological Institute Russian

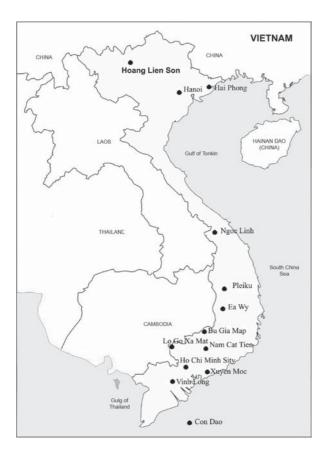


Figure 1. Original sample locations. See Table 1 for detailed information.

R. tanezumi

7 Th15

8 Sh56

9

10 Sh89

11

12 HG2

13

SVK10-03

CD1

HG7

HG8 14

Academy of Sciences, Saint-Petersburg, Russia (ZIN), or comprise the personal collections of the authors (Tab. 1).

DNA extraction, amplification and sequencing

Genomic DNA was isolated by the routine phenolchloroform method (Sambrook et al., 1989) and purified by two-fold alcohol precipitation using a DNA Purification Kit (Fermentas). A full-sized cytochrome b (Cyt b) gene (1143 bp) and/or a 680 bp portion of the subunit I cytochrome oxidase gene (COI) was amplified using the universal conservative primers H15915R (5' GGAATTCATCTCTCCGGTTTACAAGAC 3'), Cyt bRglu2L (5' CAGCATTTAACTGTGACTAAT-GAC_3'), and CytbRCb9H (5'_TACACCTAGGAG-GTCTTTAATTG 3') for the Cyt b gene and BatL 5310 (5' CCTACTCRGCCATTTTACCTATG 3') and R6036R (5'_ ACTTCTGGGTGTCCAAAGAAT-CA_3') for the COI gene (Kocher et al., 1989; Irwin et al., 1991; Robins et al., 2007). The polymerase chain reaction was conducted in 30 to 50 µl of KCl containing PCR buffer (Fermentas). The PCR mixture contained 50 mM of each dNTP, 2 mM of MgCl,, 10 to 12 pmol of each primer, 1 U Taq DNA polymerase (Fermentas) and 20 to 50 ng of DNA template. Amplification was conducted according to the following protocol: denaturation 40 s at 95°, 40 cycles 30 s long at 95°, 1 min at 48–56°, 40 s at 72°, and terminating elongation for 2 min at 72°. PCR products were purified using a DNA Purification Kit (Fermentas) following the manufactur-

FR775850

FR775851

FR775852

HO907884

HQ907885

HO907886

Accession numbers Samples information Museum No. Voucher Specimen Taxonomic number HG COI Locality Cyt b nomination nomination location 1 Th7 R. tanezumi **ZMMU** FR775845 S-184839 2 Th9 R tanezumi **ZMMU** R2 FR775846 S-184842 3 Th₁₀ R. tanezumi **ZMMU** R2 FR775847 S-184843 4 Th11 Ho Chi Minh City **ZMMU** R2 S-184844 R. tanezumi FR775848 5 Th12 **ZMMU** FR775849 S-184841 R. tanezumi R2 6 Th14 R. tanezumi **ZMMU** R2 S-184846

ZMMU

auth.coll.

ZMMU

ZMMU

auth.coll.

ZMMU

ZMMU

ZMMU

Hoang Lien Son

Hanoi

Con Dao Isl.

Hai Phong

R2

R2

R2

R2

R2

R2

R2

R2

Table 1. List of original samples.

S-188901

S-188903

S-184840

S-188105

S-184740

S-184741

S-184742

FR775804

FR775805

FR775806

FR775807

HO907882

HQ907883

Table 1 (continued).

No.		Accession numbers		Museum				
	Specimen nomination	Taxonomic nomination	Locality	Voucher location	HG	Cyt b	COI	number
15	K17	R. tiomanicus	Nam Cat Tien	auth.coll.	R3		FR775822	S-188904
16	K21	R. tiomanicus		auth.coll.	R3	FR775853	FR775808	
17	K67	R. tiomanicus		ZMMU	R3	FR775854	FR775809	S-186948
18	K89	R. tiomanicus		ZMMU	R3	FR775855		S-184848
19	F27	R. tiomanicus		auth.coll.	R3		FR775810	
20	91165nc	R. tiomanicus	Ngoc Linh	ZIN	R3		FR775811	91165
21	Th16	R. tiomanicus	HCM City	ZMMU	R3	FR775856		S-184849
22	SM1	R. tiomanicus	Xuyen Moc	ZMMU	R3	FR775857		S-188905
23	Z22	R. tiomanicus	Pleiku	auth.coll.	R3	FR775858		
24	Z37	R. tiomanicus		auth.coll.	R3	FR775859		
25	Lo25	R. tiomanicus	Lo Go Xa Mat	ZMMU	R3	FR775860		S-188908
26	Lo37	R. tiomanicus		ZMMU	R3	FR775861		S-188909
27	VL1	R. tiomanicus	Vinh Long	ZMMU	R3		FR775812	S-188910
28	VL5	R. tiomanicus		ZMMU	R3	FR775862	FR775813	S-188911
29	VL6	R. tiomanicus		ZMMU	R3		FR775814	S-188912
30	VL7	R. tiomanicus		ZMMU	R3	FR775863	FR775815	S-188913
31	VL8	R. tiomanicus		ZMMU	R3	FR775864	FR775816	S-188914
32	VL9	R. tiomanicus		ZMMU	R3		FR775817	S-188915
33	VL10	R. tiomanicus		ZMMU	R3		FR775818	S-188916
34	VL26	R. tiomanicus		ZMMU	R3	FR775865	FR775819	S-188917
35	VL42	R. tiomanicus		auth.coll.	R3		FR775820	
36	CD37c	R. tiomanicus	Con Dao Isl.	ZMMU	R3		FR775821	S-188918
37	R1	R. andamanensis	Nam Cat Tien	auth.coll.	R7	FR775866		
38	R3	R. andamanensis		auth.coll.	R7	FR775867		
39	K79	R. andamanensis		ZMMU	R7	FR775868		S-188919
40	K90	R. andamanensis		ZMMU	R7	FR775869		S-184833
41	BZM1	R. andamanensis	Bu Gia Map	ZMMU	R7	FR775870		S-188920
42	BZM3	R. andamanensis		ZMMU	R7	FR775871		S-188921
43	BZM4	R. andamanensis		auth.coll.	R7	FR775872		
44	BZM7	R. andamanensis		ZMMU	R7	FR775873		S-188922
45	BZM8	R. andamanensis		ZMMU	R7	FR775874		S-184832
46	VL22	R. argentiventer	Vinh Long	ZMMU	R6		FR775823	S-188923
47	VL23	R. argentiventer		ZMMU	R6		FR775824	S-188924
48	VL25	R. argentiventer		ZMMU	R6	FR775875	FR775825	S-188925
49	VL27	R. argentiventer		ZMMU	R6	FR775876	FR775826	S-188926
50	VL28	R. argentiventer		ZMMU	R6		FR775827	S-188927
51	VL29	R. argentiventer		auth.coll.	R6		FR775828	
52		R. argentiventer		ZMMU	R6		FR775829	S-188928
53		R. argentiventer		ZMMU	R6	FR775877	FR775830	S-188929

Table 1 (continued).

		Accession numbers		Museum				
No.	Specimen nomination	Taxonomic nomination	Locality	Voucher location	HG	Cyt b	COI	number
54	VL34	R. argentiventer	Vinh Long	ZMMU	R6	FR775878	FR775831	S-188930
55	VL35	R. argentiventer		ZMMU	R6	FR775879	FR775832	S-188931
56	VL36	R. argentiventer		ZMMU	R6	FR775880	FR775833	S-188932
57	VL39	R. argentiventer		auth.coll.	R6	FR775881	FR775834	
58	VL41	R. argentiventer		auth.coll.	R6	FR775882	FR775835	
59	D29	R. nitidus	Ea Wy	ZMMU	R10	FR775883		S-188933
60	Z40	R. nitidus	Pleiku	ZMMU	R10	FR775884		S-188934
61	VL3	R. exulans	- Vinh Long	ZMMU	R8		FR775836	S-188935
62	VL4	R. exulans		ZMMU	R8		FR775837	S-188936
63	VL12	R. exulans		ZMMU	R8		FR775838	S-188937
64	VL13	R. exulans		auth.coll.	R8		FR775839	
65	VL14	R. exulans		auth.coll.	R8		FR775840	
66	VL15	R. exulans		auth.coll.	R8		FR775841	
67	VL40	R. exulans		auth.coll.	R8	FR775885	FR775842	
68	VL43	R. exulans		auth.coll.	R8	FR775886	FR775843	
69	Th1	R. norvegicus	Ho Chi Minh City	auth.coll.	R9	FR775887		
70	Th3	R. norvegicus		ZMMU	R9	FR775888		S-184835
71	Th4	R. norvegicus		ZMMU	R9	FR775889		S-184836
72	Th8	R. norvegicus		ZMMU	R9	FR775890		S-188938
73	K76	B. indica	Nam Cat Tien	ZMMU		FR775891		S-188939
74	VL24	B. indica	Vinh Long	ZMMU			FR775844	S-188940

er's protocol and sequenced by Applied Biosystems 3130 Genetic Analyzer using an ABI PRISM BigDye Terminator Cycle Sequencing Ready Reaction Kit for both nucleotide chains, using the same primers as for the initial PCR.

All Cyt b and COI gene sequences that were obtained and used for comparative analysis of genetic distances and phylogenetic groups delimitation were deposited into EMBL databank (FR775804–FR775891, Tab. 1). A number of gene sequences and species used as comparative material were cited by Suzuki et al. (2000, 2003; AB033701-AB033702, AB096841), Chinen et al. (2005; AB211039-AB211053), Schlick et al. (2006; DQ 673907-DQ673914), Robins et al. (2007; EF186409-EF186512, EF186524-EF186528, F186559-EF186561, EF186605–EF186627), Truong et al. (2009; AB355899– AB355901), Pages et al. (2010; HM217362-HM 217596), Tollenaere et al. (2010; GQ891569-GQ 891608), and Bastos et al. (2011; DQ439816-DQ 439864). Totally, we studied 227 specimens with geographic locations for the Cyt b gene and 178 specimens for COI, including original 49 Cyt b and 43 COI sequences from 72 individuals of Rattus from Vietnam. We also used one specimen of *M. musculus* (J01420) as a distant out-group and two specimens of *Bandicota indica* from Vietnam as neighboring out-groups (Tab. 1).

Sequences alignment and phylogenetic analysis

Sequence alignment was carried out using the software program BioEdit (Hall, 1999). The phylogenetic trees were constructed using minimum evolution (ME) and neighbor-joining (NJ) methods. Genetic distances were calculated by the Tamura 3-parameter model (T3P) for all types of substitutions (Ts+Tr) using MEGA 3.1 software (Kumar *et al.*, 2004). The bootstrap support level for nodes was calculated by 1000 replications. Final correspondence between specific level haplogroups/species and any morphologically delimitated species was performed in agreement with the genetic species concept (Bradley & Baker, 2001; Baker & Bradley, 2006; Pages *et al.*, 2010). All COI gene sequences are also included in our collaborative project named Indochinese Muridae (ICMBA) at Barcoding of

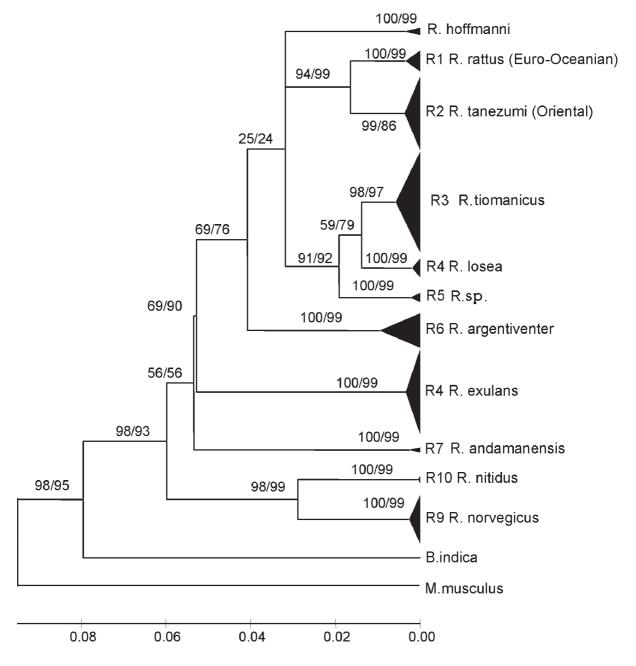


Figure 2. Phylogenetic relationships within *Rattus* species inhabiting the Sunda Shelf region; ME tree. Distal branches are shown compressed. Species nomination generally follows Robins *et al.* (2007) but see discussion below. Bootstrap values for the Cyt *b*/COI genes are shown above branches. Intergroup divergence scale is at the bottom.

Life System, BOLD (Savolainen *et al.*, 2005; www. barcodinglife.com) and have been used for interspecific comparisons within the BOLD system.

Results

Genetic phylogroup compositions

Nine specific level haplogroups of the *Rattus* spp. can be recognized among the data set from Sundaland, with seven inhabiting Vietnam (Fig. 2). Five of them

are closely related species of the *Rattus* group, with hardly resolved interrelations within this subcluster. Between group means genetic distances, (d $_{\rm bg}$) for Cyt b is 0.038–0.055; for COI is 0.028–0.069). The topologies of the trees that were constructed based on Cyt b and COI sequences using both the NJ and ME methods were almost identical, with only negligible differences in bootstrap values for some of the distal clades. Thus, we present only a generalized tree, shown in Figure 2. Intra- and intergroup genetic distances (d $_{\rm bg}$, d $_{\rm wg}$, Tajima 3-parameter, T3P) are shown in Table 2.

Species n 30 0.005 0.005 0.007 0.007 0.006 0.010 0.010 0.008 0.011 0.011 0.011 R1, R. rattus 9 0.004 0.007 0.012 0.011 0.0110.011 0.012 0.015 0.016 0.016 0.019 61 0.041 0.008 0.007 0.010 0.009 0.012 0.013 0.009 0.007 0.010 0.012 R2, R. tanezumi 29 0.033 0.005 0.010 0.009 0.010 0.011 0.011 0.014 0.015 0.015 0.018 41 0.057 0.065 0.011 0.006 0.006 0.009 0.010 0.009 0.011 0.011 0.011 R3, R. tiomanicus* 44 0.075 0.062 0.005 0.007 0.008 0.010 0.011 0.013 0.014 0.015 0.018 0.068 8 0.062 0.038 0.003 0.006 0.009 0.010 0.009 0.011 0.010 0.011 R4, R. losea 8 0.067 0.057 0.028 0.002 0.008 0.010 0.012 0.013 0.015 0.016 0.018 4 0.040 0.053 0.041 0.041 0.005 0.008 0.010 0.010 0.012 0.012 0.011 R5, R. sp.* 5 0.069 0.037 0.003 0.009 0.014 0.014 0.015 0.062 0.039 0.012 0.017 3 0.074 0.080 0.059 0.055 0.012 0.013 0.013 0.066 0.004 0.011 0.012 R. hoffmanni 3 0.068 0.067 0.064 0.060 0.058 0.005 0.012 0.014 0.015 0.014 0.017 11 0.096 0.101 0.101 0.095 0.085 0.078 0.008 0.010 0.012 0.013 0.012 R6, R. argentiventer 15 0.089 0.081 0.080 0.082 0.084 0.083 0.014 0.015 0.015 0.018 0.014 11 0.078 0.089 0.088 0.088 0.079 0.092 0.101 0.012 0.011 0.012 0.012 R7, R. andamanens 2 0.109 0.102 0.097 0.090 0.095 0.096 0.005 0.017 0.016 0.018 0.110 37 0.101 0.109 0.097 0.100 0.093 0.103 0.115 0.098 0.005 0.014 0.013 R8, R. exulans 36 0.121 0.110 0.096 0.098 0.101 0.108 0.1210.128 0.003 0.016 0.016 0.008 16 0.120 0.132 0.113 0.112 0.101 0.111 0.133 0.130 0.137 0.004 R9, R. norvegicus 21 0.119 0.114 0.111 0.123 0.113 0.110 0.124 0.119 0.128 0.003 0.010 5 0.132 0.139 0.125 0.127 0.105 0.105 0.143 0.127 0.135 0.060 0.007 R10, R. nitidus 3 0.121 0.058 0.144 0.127 0.128 0.1360.123 0.130 0.144 0.134 0.001

Table 2. Mean genetic distances among *Rattus* spp. from the Sunda Shelf region.

Note: The distances calculated based on the Cyt b gene are in the upper lineage, and the distances calculated based on the COI gene are in the lower lineage. Between-group means (d_{bg}) are in the bottom left, and standard errors (S.E.) are in the top right, within group means (d_{wg}) in the diagonal.

* See discussion about probable proper taxonomic name for this species in the text.

For full integrity of background and to cover all currently characterized genetic species of the Rattus group in the Asia-Pacific region, we also brought the sequences treated as R. tiomanicus Miller, 1900 (R5) and R. hoffmanni Matschie, 1901 by Robins et al. (2007, 2010) to our study, which rise the number of species studied to 11. Haplogroups nominations (R1– R10) follow Pages et al. (2010). Because the majority of our material represented Southern Vietnam, R. losea (R4), the species typical in central and northern Indochina, is lacking in our samples. Only few information is currently available on the general features of the species including its actual geographic ranges and taxonomical limits. Even so, there are not any doubts that this species is present in the fauna of the country. Another species, which morphologically close to *R*. losea, R. osgoodi Musser et Newcomb, 1985 (Musser & Newcomb, 1985), describing from Langbian Peak in Dalat Plateau, is still not characterized genetically. This species is apparently present in the fauna of Vietnam, but there are no doubts about the point that any of the genetic lineages examined in the paper actually represent *R. osgoodi*. Thus, formal diversity in the genus *Rattus* is shown to be within the previously established limits (Musser & Carleton, 2005), but to obtain a better estimation of taxonomic composition, an analysis of correspondence of genetic lineages to morphological species is needed.

Discussion

Analysis of phylogroups geographical distributions

Using an analysis of capture site locations and phylogenetic relationships between genetic groups, the conclusion can be drawn which may not completely

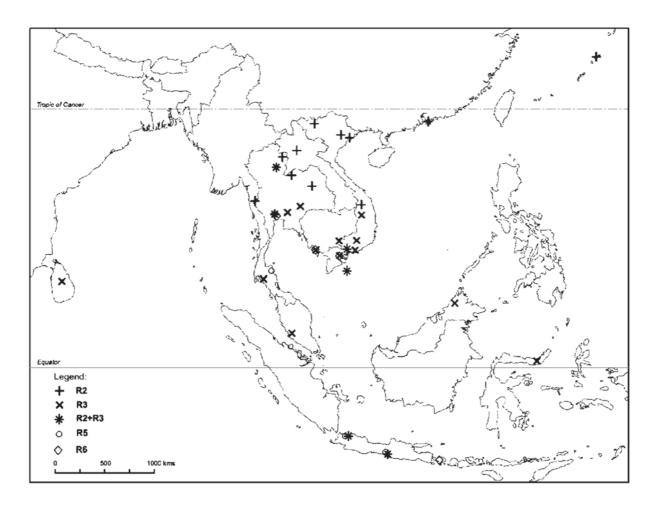


Figure 3. Distribution of some specific level haplogroups/species of the genus *Rattus* in the Sunda Shelf region. Haplogroup nominations follow Pages *et al.* (2010). Site localities for R3 in Kalimantan and R6 in Java presented by morphological data.

agree with the generally accepted taxonomic concept of the genus *Rattus* (Musser & Carleton, 2005; Pavlinov, 2005). Under this conception, *R. argentiventer* Robinson and Kloss, 1916 and *R. tiomanicus* are restricted in its northern range by Isthmus of Kra. It can be seen in Figure 3 that no less than three genetic groups (R3, R5, R6), probably corresponding to *R. tiomanicus*, *R. argentiventer* and one other unclearly nominated species, represent a wide distribution both to the north and to the south from the most significant zoogeographic limit in the region, the Isthmus of Kra. Thus, it may be stated that there is genetic uniformity within the whole series of continental and Sundaic species instead of insulation and specific segregation of forms in the area of the Sunda Shelf, as suggested earlier.

Based on available genetic data, it is evident that the most common species in continental Indochina (Thailand, Cambodia, and Vietnam), generally referred to as *R. rattus* Linnaeus, 1758 or *R. tanezumi*, in fact could not be regarded as either of these species. This species treated as *R. diardi* in Robins *et al.* (2007) and Pages *et al.* (2010). However, the specific name *diardi* is currently excluded from a list of valid synonyms for any of

the generally accepted *Rattus* species (Musser & Carleton, 2005). Moreover, there have been no reports of *diardi* spreading to the north beyond the Isthmus of Kra, though this semisynanthropic *rattus*-group species widely distributed on the Malay Peninsula, on some islands of the Sunda Archipelago (Java, Sulawesi) and well beyond the region right up to Sri Lanka. In Southern Vietnam, the rats preferably inhabit vegetable gardens, rice fields, grasslands and shrubs near human settlements; sometimes, though rarely, they can be caught within human houses.

It have not been noticed previously on the range of *R. tiomanicus* (R5, follows Robins *et al.*, 2007, Pages *et al.*, 2010) spreading northward from the previously described zoogeographical limit, which, as has been shown by above mentioned authors, spreads well to the north on peninsular Thailand and Cambodia. However, we failed to record this haplogroup in Vietnam. The genetic lineage R6, which appears to be *R. argentiventer*, is also shown to be distributed well beyond the Isthmus of Kra and has been recorded both in Southern Cambodia (Pages *et al.*, 2010) and in the Mekong delta (our data).

When we combined all of the material for the *rattus* group available in GenBank (more than 300 specimens) with our original data from Vietnam, we failed to find any individuals belonging to haplogroup R1, considered as R. rattus in the Indochina and Sundaland areas. This species is recorded in the region only from New Zealand, Polynesia and Japan. It is should be mentioned that R. rattus and R. tanezumi could hybridize and produce fertile hybrids (Chinen et al., 2005), but any genetic traces of R. rattus are still absent in the region. In spite of the fact the absence cannot be logically approved, it is obvious that if the European black rat R. rattus actually forms any stable populations anywhere in Indochina (most likely in ports, airports or in warehouses and trans-shipment points), their distribution here actually represents sporadic foci or enclaves, expansions from which are apparently hampered by competition with indigenous species, mainly R. tanezumi, R. losea and R. argentiventer. Detailed investigation of such types of habitats was beyond of the scope of our study, so we failed to find any of such enclaves. However, to avoid confusion, it is recommended to be very prudent when using the specific name R. rattus anywhere in Indochina.

Therefore, based on available data, we can conclude the lack of true R. rattus from the fauna of Indochina and the wide distributions of many species considered to be endemic to the Sundaic and Malayan zoogeographical subregions. If the former phenomenon can be explained naturally, the latter may have arisen from misidentification of original samples and erroneous taxonomic name usage. Regardless, based on the obvious genetic uniformity of whole series of forms in the regions both to the north and south from the Isthmus of Kra, questions arise regarding the taxonomic status of these forms and the validity of their scientific names. Unfortunately, earlier authors (Robins et al., 2007; Pages et al., 2010) were guided only by the authors' identifications of the museum samples from a label and by a principle of locality to decide on the appropriate taxonomic names for revealed genetic clades; the morphological characters of specific museum specimens were not examined. We acknowledge the utility and higher resolution of comprehensive genetic approach for the purposes of species diagnosis, but it is necessary to emphasize that for such complex group as the genus Rattus in the Asia Pacific region, it is essential to compare any genetic data with the original morphological and museum material to determine the correct taxonomic names for the samples.

Contribution on taxonomical status and nomenclature for recorded genetic lineages

We attempted to makes clearer what exactly taxonomic names may be correct or, at least, as most as appropriate to attribute to genetic lineages/species discovered using not only genetic but also morphological data (skins, skulls, carcasses and photo images along

with a personal observations from live animals) for our Vietnamese specimens, many of which proved to be genetically identical to many of the forms discussed in Robins *et al.* (2007, 2010) and Pages *et al.* (2010) (R2, R3, R6, R7, R8, R9, and R10). Comparison of the general appearance and distinctive morphological features of the skull for revealed specific level phylogroups, allowing us to assume its association with some forms and species which have been described early only morphologically.

From seven Vietnamese genetic lineages mentioned above, the attribution for four haplogroups, namely R2 (R. tanezumi), R8 (R. exulans Peale, 1848), R9 (R. norvegicus Berkenhout, 1769), and R10 (R. nitidus Hodgson, 1845), was not difficult to determine based on their obvious morphological peculiarities. Likewise, the attribution of haplogroups R1 (R. rattus) and R2 (R. tanezumi) was recently performed by Chinen et al. (2005) based on karyologically confirmed specimens representing both of the species captured in several sites in Japan, including one place in close vicinity to the possible terra typica for R. tanezumi, near Nagasaki, Kyushu Island (see Jones & Johnson, 1965). Three other species also bear may be not so prominent, but clear enough morphological appearances and their specific diagnosis can be made based on comparative analysis of general and skull morphology. All of them were recorded in fauna of Vietnam; they also demonstrate well-separated, distinct clades on a phylogenetic tree. Nevertheless, there has been long term discussion regarding the identities of other species, R. argentiventer, R. andamanensis Blyth, 1860 (syn. R. sikkimensis/R. koratensis/R. remotus), R. diardi and R. tiomanicus and on the possibility of their correspondences to any of the genetic lineages R3, R5, R6, and R7. It should be noted that, in general case, there is not any strong needs to perform any profound and spectacular statistical analysis to distinguish some peculiar traits which characteristic for the species at their skull structure. Fig. 4 shows photos of the skulls for representatives of the genetic lineages R2, R3, R6 and R7. To compare, we also present reproductions of the skulls (including some types) of different forms and species of genus Rattus from Indochina and the Sunda Archipelago from reviews of Musser (Musser & Califia, 1982; Musser & Newcomb, 1985; Musser, 1986). Images of the skull for some *Rattus* species are also available on the Internet site of the Field Museum of Natural History, Chicago, USA (http://www.fieldmuseum.org).

It is the most simple to establish an attribution of group R6 to species *R. argentiventer* using peculiarities of its general appearance (rust-colored back and entire upper part of body, short ears, dull ochreous spot in front of the ear, light gray, silver-colored belly, dark hind feet; short, broad and lyre-shaped narrowing ahead of the rostrum and very broad and spectacularly formed incisive foramina stretching to the fore edge of the first molars, Fig. 4A, H). These rats inhabiting rice fields in the Mekong delta undoubtedly belong to *R. argentiven*-

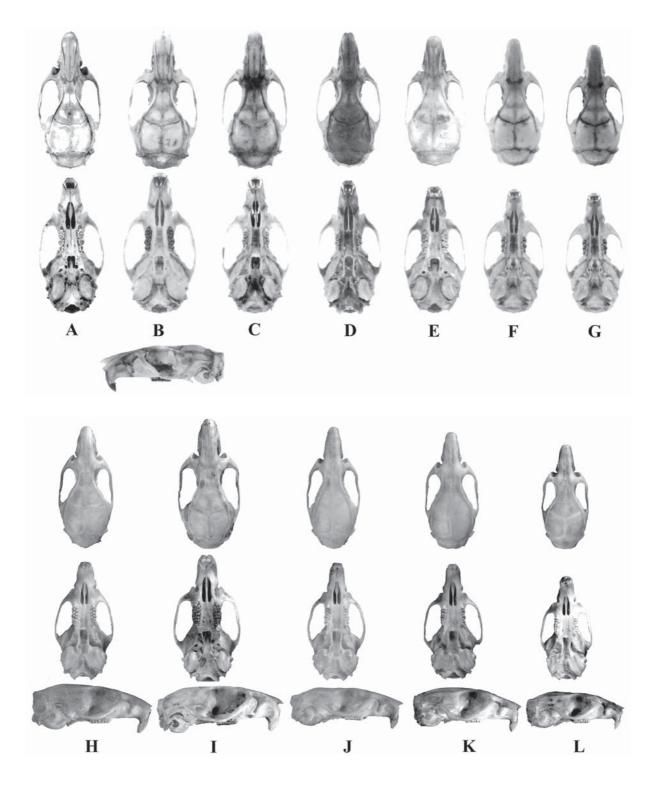


Figure 4. Skulls of adult Rattus from the Sunda Shelf (after Musser, 1986; Musser & Califia, 1982; Musser & Newcomb, 1985) and Southern Vietnam (original photo): À — R. argentiventer (AMNH 107543, Bali Island); B — R. sikkimensis/R. andamanensis (USNM 533456, Thailand); C — R. diardi (AMNH 250104, Java); D — R. diardi (USNM 292689, Gunung Kinabalu, Sabah); E — R. tiomanicus roquei (AMNH 250114, Java); F — R. tiomanicus sabae (USNM 292789); G — R. tiomanicus (USNM 292672, Sabah); H — R. argentiventer, R6 (VL35, Vinh Long); I — R. andamanensis, R7 (BZM1, Bu Gia Map); J — R. tiomanicus, R3 (Lo37, Lo Go Xa Mat); K — R. tiomanicus, R3 (VL26, Vinh Long); L — R. tiomanicus, R3 (SM1, Xuyen Moc).

ter (Musser & Newcomb, 1983; Corbet & Hill, 1992). Thus, we can confirm a correct taxonomic name for this haplogroup and approved the distribution of the species well to the north from the earliest presumed natural area (Malay Peninsula southward of the Isthmus of Kra), at least as far as the Mekong delta.

Comparison of craniometrical peculiarities of our original specimens with descriptions of morphology and images of skulls presented in the literature (Fig. 4) allowed us to attribute haplogroup R7 to Rattus sikki*mensis*, currently synonymized with *R. andamanensis*. The skull structure of these forms is very similar (Fig. 4B, I). The coarse structure of the skull with profound angular shelves and ridges, very spectacular shaped, well-elongated, narrow and tapering into extremities incisive foramina, and especially small, low and visually appreciable angular bulla (which is most spectacular on side view), are all characters composing distinctive features of the species. The rat is an entirely wild (nonsynanthropic) species that has been caught in Nam Cat Tien National Park and Bu Gia Map Nature Reserve in matured non- or lightly-disturbed dipterocarp forests. It is exactly the same genetic lineage listed for Western Thailand reported by Pages et al. (2010). That is fairly large rat with a chestnut-colored or dull brown upper part and a fairly coarse coat, bearing long dark brown or black guard hairs along the back, which penetrate the main hair coat and project more than 1 cm beyond. In Nam Cat Tien, the species has a perceptible yellowish hue or even obvious yellow belly with fairly well demarcated borders between the coloration of upper and lower parts of body, whereas specimens from Bu Gia Map are more consistent with descriptions of the morphae koratensis/remotus in Corbet & Hill (1992) in their appearance and have a dirty-whitish or creamy abdomen. The hind feet are broad, and the dorsal sides are also uniformly yellowish, ochreous-colored or bear a broad grayish band. In contrast to the most other Rattus species, the tail is covered to the very tip by short tough brownish hairs, with the dorsal side colored appreciably lighter, with a visible limit of colors transition. Thus, we have to agree again with the accuracy of the taxonomic species name for haplogroup R7 as corresponding to R. andamanensis and approve distribution of the species to East Indochina. It is worth emphasizing that the species is very similar in its general appearance with many of the insular morphae of Rattus traditionally treated as R. diardi (Fig. 4C, D) and now treated as R. rattus, R. tiomanicus or R. tanezumi. However, there is still not any available genetic data for haplogroup R7 from anywhere beyond continental Indochina. The fact that the species was described from the Andaman Islands allows us to assume its distribution (or the existence of any sister or sibling species) in the Sunda Shelf islands. If this species is actually inhabits these locations, the taxonomical name diardi may be applied to them. However, R. andamanensis comprises an independent clade in the genus that is genetically well-segregated from species of the rattus group

(*R.rattus*, *R. tanezumi*, *R. losea*, *R. tiomanicus*; haplogroups R1–R4 and *R. hoffmanni*), so it is hardly if any correct to place it among this group of species.

However, the conclusions drawn above vary from and even contradict with the conclusions of Robins et al. (2007, 2010) and Pages et al. (2010) in whether the name diardi should be attributed to haplogroups R3. By comparing external appearance (white belly, sleek, soft and silky hair coat, a tail much longer than body length) and skull structure (Fig. 4 I-G and J-L) where an appreciably larger bulla can be seen, oblong shaped rostrum, much less pronounced bone ridges, slender, even fragile zygomatic arches, more long and broad row of molars. It can be clearly recognized that the species inhabiting south of Vietnam, Thailand and Cambodia and bearing haplotype R3 is a member of the rattus group but certainly not diardi, the morph to which rats having a appreciably different appearance (much larger and generally darker colored with coarse hair coat and invariably darker, brownish-yellow, buff or dark gray abdomen) have generally been attributed.

As shown in Fig. 3, the haplogroup R3 is independent, high-supported clade and cannot be ascribed to any other species of the rattus group, R. rattus (R1), R. tanezumi (R2) or R. losea (R4). The external appearance, skull structure and especially their natural area (wide distribution of the haplogroup R3 on many Sunda Archipelago islands, to Java and even Sulawesi) permit us to assume the attribution of the morph to R. tiomanicus. It is exactly this rat species from the rattus group (smaller in size, with white belly and sleek and silky hair coat, sometimes erroneously identified and treated under the name R. tanezumi) for which this name was earlier being used (Musser & Califia, 1982; Musser & Newcomb, 1985; Musser, 1986; Corbet & Hill, 1992; Musser & Carleton, 2005). Thus, the facts allow us to assume that the most abundant and most widely distributed species of the rattus group in Southern Vietnam is the species also distributed throughout most of Thailand, Cambodia, the Malay Peninsula, the islands of Java and Sulawesi and even Sri Lanka (Pages et al., 2010; Robins et al., 2007, 2010). The taxon should be named as R. tiomanicus.

This species has some distinctive features in their external morphology that can help separate it from conjointly distributed R. argentiventer or R. tanezumi. To distinguish it from the former, it has a clear white belly and dull, brownish, rather than reddish, coloration of its backside and a light grayish strip instead of homogenous gray coloration on the dorsal side of the feet. In contrast to some individuals of R. tanezumi, which also have a white abdomen, R. tiomanicus bears a widely-spread rather than closely-approximating caudal pair of mammilla and perceptible (especially for adult males) long guard hair coming out of the main coat nearly 1 cm. The last feature is often noted as the distinctive character for R. remotus/R. andamanensis (Marshall, 1977), but the latter is completely wild forest species having a distinctly different hair coat texture, tail downiness, belly coloration and a specific skull compared to any species of *rattus* group.

If haplogroup R3 may be attributed to R. tiomanicus, the question still exists on a proper taxonomic name for haplogroup R5, which inhabits both Peninsular Thailand (Pages et al., 2010), and Jakarta and the Cibodas Forests in Java, Indonesia (Robins et al., 2007). Unfortunately, these authors did not discuss the external appearance, so nothing is known on the morphological peculiarities of this rats. We failed to find any haplotypes of the R5 group in Vietnam and cannot reasonably speculate about their taxonomical attribution. However, it may well be Rattus anandalei Bonhote, 1903 can be used as one of possible name for it which is known to inhabit the Malay Peninsula, Singapore and neighboring islands (Musser & Newcomb, 1983). The species is also supposed to belong to the rattus group but shows some specific characters in their morphology preventing it to be drawn in with any other morphae from this study.

It is also worth to note that the fact the *R. tiomanicus* inhabiting continental Indochina may well pose a question about privilege of this taxonomic name as the oldest and hence general synonym. The species *R. germani* Milne-Edwards, 1872 was described from Con Son Island (Con Dao Archipelago, Southern Vietnam) and has priority over *R. tiomanicus* Miller, 1900 from Tioman Island (Malaysia). However, we found that the Con Son Island is actually inhabited by representatives of two haplogroups/species, R2 *R. tanezumi* and R3 *R. tiomanicus* (Tab. 1). With this fact being the case and bearing in mind that the initial description was not explicit and lacking the type material, it is hardly possible to argue which of these species the description actually mentioned.

Finally, we would like to express a desire that subsequent papers on the phylogeny and phylogeography of the genus Rattus be based on the established numeration of phylogenetic groups following Robins et al. (2007) and Pages et al. (2010) with our amendment: R1 (R. rattus, Euro-Oceanian type), R2 (R. tanezumi, Oriental type), R3 (R. tiomanicus, Sundaic type), R4, (R. losea), R5 (perhaps R. anandalei, needs more precision), R6 (R. argentiventer), R7 (R. andamanensis), R8 (R. exulans), R9 (R. norvegicus) and R10 (R. nitidus). The numeration may be developed and extended by applying indexes for newly discovered genetic lineages. The system of nomination allows more certainty in established taxonomy for a feature and finally attaining full comparability of the data obtained by independent researchers.

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